## Quantifying the response of photosynthesis to changes in leaf nitrogen content and leaf mass per area in plants grown under atmospheric CO<sub>2</sub> enrichment

# A. G. PETERSON,<sup>1,4</sup> J. T. BALL,<sup>1</sup> Y. LUO,<sup>1</sup> C. B. FIELD,<sup>2</sup> P. S. CURTIS<sup>3</sup>, K. L. GRIFFIN,<sup>4</sup> C. A. GUNDERSON,<sup>5</sup> R. J. NORBY,<sup>5</sup> D. T. TISSUE,<sup>6</sup> M. FORSTREUTER,<sup>7</sup> A. REY,<sup>8</sup> C. S. VOGEL<sup>9</sup> & CMEAL PARTICIPANTS<sup>10</sup>

<sup>1</sup>Biological Sciences Center, Desert Research Institute, PO Box 60220, Reno, NV, 89506, USA, <sup>2</sup> Carnegie Institution of Washington, Department of Plant Biology, 260 Panama Street, Stanford, CA 94305, USA, <sup>3</sup> Department of Plant Biology, Ohio State University 1735 Neil Ave., Columbus, OH 43210, USA, <sup>4</sup> Earth and Environmental Sciences, Lamont-Doherty Earth Observatory, Columbia University, 61 Route 9 W, Palisades, NY 10964, USA, <sup>5</sup>Environmental Sciences Division, Oak Ridge National Laboratory, PO Box 2008, Oak Ridge, TN 37831, USA, <sup>6</sup> Department of Biology, Flint and Main Street, Texas Tech University, Lubbock, TX 79409, USA, <sup>7</sup> Technical University Berlin, Institute of Ecology, Koenigin-Luise-Str.22, D-14195 Berlin, Germany, <sup>8</sup> IERM, King's Buildings, Edinburgh University, Mayfield Road, Edinburgh, EH9 3JU, UK, <sup>9</sup> University of Michigan Biological Station, 9008 Biological Rd, Pellston MI, 49769, USA and <sup>10</sup> CMEAL (for details see Appendix)

## ABSTRACT

Previous modelling exercises and conceptual arguments have predicted that a reduction in biochemical capacity for photosynthesis (Aarea) at elevated CO2 may be compensated by an increase in mesophyll tissue growth if the total amount of photosynthetic machinery per unit leaf area is maintained (i.e. morphological upregulation). The model prediction was based on modelling photosynthesis as a function of leaf N per unit leaf area  $(N_{area})$ , where  $N_{area} =$  $N_{\text{mass}} \times LMA$ . Here,  $N_{\text{mass}}$  is percentage leaf N and is used to estimate biochemical capacity and LMA is leaf mass per unit leaf area and is an index of leaf morphology. To assess the relative importance of changes in biochemical capacity versus leaf morphology we need to control for multiple correlations that are known, or that are likely to exist between  $CO_2$  concentration,  $N_{area}$ ,  $N_{mass}$ , LMA and  $A_{area}$ . Although this is impractical experimentally, we can control for these correlations statistically using systems of linear multipleregression equations. We developed a linear model to partition the response of Aarea to elevated CO2 into components representing the independent and interactive effects of changes in indexes of biochemical capacity, leaf morphology and CO<sub>2</sub> limitation of photosynthesis. The model was fitted to data from three pine and seven deciduous tree species grown in separate chamber-based field experiments. Photosynthetic enhancement at elevated CO<sub>2</sub> due to morphological upregulation was negligible for most species. The response of  $A_{\text{area}}$  in these species was dominated by the reduction in CO<sub>2</sub> limitation occurring at higher CO<sub>2</sub> concentration. However, some species displayed a significant reduction in potential photosynthesis at elevated CO<sub>2</sub> due to an increase in *LMA* that was independent of any changes in  $N_{\text{area}}$ . This morphologically based inhibition of  $A_{\text{area}}$  combined additively with a reduction in biochemical capacity to significantly offset the direct enhancement of  $A_{\text{area}}$  caused by reduced CO<sub>2</sub> limitation in two species. This offset was 100% for *Acer rubrum*, resulting in no net effect of elevated CO<sub>2</sub> on  $A_{\text{area}}$  for this species, and 44% for *Betula pendula*. This analysis shows that interactions between biochemical and morphological responses to elevated CO<sub>2</sub> can have important effects on photosynthesis.

*Key-words*: carbon dioxide; leaf nitrogen; photosynthesis; structural equation model.

## INTRODUCTION

Leaf-level photosynthesis  $(A_{area})$  is often enhanced in plants grown under long-term exposure to elevated levels of atmospheric CO<sub>2</sub> (Gunderson & Wullschleger 1994; Curtis 1996; Drake, Gonzàlez-Meler & Long 1997). This enhancement is caused by an increase in the rate of carboxylation of ribulose-1,5-bisphosphate (RuBP) catalyzed by RuBP carboxylase/oxygenase (Rubisco) (Woodrow & Berry 1988). There are at least two fundamentally different ways in which the rate of carboxylation per unit leaf area can be modified by elevated CO<sub>2</sub>. The first way, which we refer to as a 'direct' effect of CO<sub>2</sub>, involves the biochemistry of photosynthesis. This direct effect consists of (i) a reduction in substrate limitation of Rubisco catalysis (Farquhar, von Caemmerer & Berry 1980) (ii) competitive reduction of RuBP oxygenation (Farguhar et al. 1980), and (iii) any adjustments in the photosynthetic apparatus (from

Correspondence: Andrew G. Peterson, Earth and Environmental Sciences, Lamont-Doherty Earth Observatory, Columbia University, 61 Route 9 W, Palisades, NY 10964, USA. Fax: USA + (914) 365–8150; e-mail: peterson@LDEO.Columbia.edu

light capture through starch and sucrose synthesis) that alter the RuBP limitation of Rubisco (Sage, Sharkey & Seemann 1989; Sage 1990). The second way in which elevated  $CO_2$  may affect the rate of carboxylation per unit leaf area involves changes in leaf morphology and anatomy (Gunderson & Wullschleger 1994; Luo, Field & Mooney 1994). These may include changes in mesophyll cell number per unit leaf area (Vu, Allen & Bowes 1989), mesophyll thickness (Sims, Seemann & Luo 1998), carbohydrate concentration (Stitt 1991) and leaf mass per unit area (*LMA*) (Curtis 1996).

During short-term exposure to elevated CO<sub>2</sub>, photosynthesis is frequently limited by the capacity to regenerate RuBP. In the longer-term, the amount or activity of Rubisco may decrease, thus balancing the reduction in RuBP regeneration (Sage 1990; Stitt 1991; Gunderson & Wullschleger 1994; Drake et al. 1997). The effect of this down-regulation of the amount or activity of Rubisco on Aarea may be offset if it is associated with an increase in mesophyll tissue such that the amount of photosynthetic apparatus per unit leaf area is maintained (Radoglou & Jarvis 1990; Luo et al. 1994). Alternatively, other changes in leaf morphology or anatomy associated with an increase in LMA may potentially reduce  $A_{\text{area}}$  since photosynthesis and LMA are negatively correlated at ambient CO2 (Reich, Walters & Ellsworth 1997; Peterson et al. 1999). This negative correlation is independent of any effect LMA may have on  $N_{\text{area}}$  or  $N_{\text{mass}}$  (Peterson *et al.* 1999) and may be due to a reduction in N allocation to photosynthetic versus nonphotosynthetic compounds (Evans 1989), to greater allocation of biomass to structural versus photosynthetic components (Vitousek, Field & Mantson 1990; Lloyd et al. 1992), to reduced light penetration through the various layers of leaf tissue (Terashima & Hirosaka 1995), or to greater limitations to internal diffusion (Parkhurst 1994; Pachepsky et al. 1997).

While biochemical aspects of photosynthetic enhancement at elevated  $CO_2$  have been studied extensively, less emphasis has been placed on understanding how those enhancements interact with changes in leaf morphology and anatomy. In an important advancement, Pachepsky & Acock (1996) developed a two-dimensional model of leaflevel photosynthesis that demonstrates how leaf anatomy can have an important effect on photosynthesis. However, the general utility of this model is limited because it requires detailed maps of mesophyll cell location and information on stomatal area and density to parameterize it. Luo et al. (1994) developed the Photosynthetic Acclimation to  $CO_2$  (PAC) model which is much less data-intensive and which suggests that interactions between biochemical and morphological responses to elevated CO2 can have important effects on leaf-level photosynthesis. The PAC model uses the biochemical model of Farquhar et al. (1980) to model photosynthesis as a function of nitrogen per unit leaf area  $(N_{\text{area}})$  (e.g. Harley et al. 1992). The model assumes that (i) nitrogen per unit leaf mass  $(N_{\text{mass}})$  reflects the biochemical capacity for photosynthesis (e.g. Körner & Miglietta 1994) since the majority of leaf N is found in the

proteins of the Calvin cycle (including Rubisco) and the thylakoid membranes (Evans 1989) and (ii) that LMA summarizes aspects of leaf morphology and anatomy such as photosynthetic biomass and non-structural carbohydrate storage. The PAC model links changes in biochemical capacity and leaf morphology through the relationship  $N_{\text{area}} = N_{\text{mass}} \times LMA$ . The model predicts that a reduction in biochemical capacity at elevated  $CO_2$  (estimated from a reduction in  $N_{\text{mass}}$ ) may be counterbalanced by morphological upregulation if an increase in LMA is relatively greater than the reduction in  $N_{\text{mass}}$ . Leaf mass per unit area has been shown to be tightly correlated with leaf thickness in soybeans grown at ambient and elevated CO<sub>2</sub> (Sims et al. 1998), but there is also substantial evidence to show that increases in LMA at elevated CO2 can be caused by increased storage of non-structural carbohydrates (e.g. Roumet *et al.* 1996). However, if both  $N_{\text{area}}$  and *LMA* increase while  $N_{\text{mass}}$  decreases, then the increase in LMA must involve additional N-bearing tissue even though N concentration decreased.

The PAC model provides valuable insights into relationships between biochemical and morphological responses to elevated  $CO_2$  by showing how net changes in  $N_{mass}$  and LMA can interact to affect photosynthesis. However, because the PAC model focuses on net changes, it does not allow us to accurately assess the relative importance of biochemical versus morphological responses. This could be achieved by experimentally controlling  $N_{\text{mass}}$  or  $N_{\text{area}}$  in addition to LMA, but this is clearly impractical. An alternative approach would be to use a simple multiple regression of  $A_{\text{area}}$  on CO<sub>2</sub> concentration,  $N_{\text{area}}$  and LMA. Since the coefficients in a multiple regression are partial derivatives, they quantify the relationship between the dependent variable  $(A_{area})$  and each independent variable when the other independent variables in the equation are held constant. This allows us to statistically control both  $N_{\text{area}}$  and LMA, and to partition the response of  $A_{\text{area}}$  among each independent variable. Using this approach, the partial regression coefficient for CO2 represents an index of CO2 limitation of photosynthesis because it quantifies the change in  $A_{\text{area}}$  per unit change in CO<sub>2</sub> when both  $N_{\text{area}}$  and LMA are held constant. Similarly, the partial regression coefficient for  $N_{\text{area}}$  represents an index of biochemical capacity per unit leaf area, and the coefficient for LMA represents an index of morphological regulation.

The multiple regression discussed above would provide useful information on the relative importance of the effects of  $CO_2$  concentration,  $N_{area}$  and LMA on photosynthesis. This model is, however, too simplistic to capture the range of interactions that are known, or that are likely to occur between these variables. To start with, both  $N_{area}$  and  $N_{mass}$ are correlated with LMA at ambient  $CO_2$  (Reich *et al.* 1994; Reich & Walters 1994; Peterson *et al.* 1999). This lack of independence may confound parameter estimation and interpretation in the simple regression model. In addition, the response of  $N_{area}$ ,  $N_{mass}$  and LMA to elevated  $CO_2$  are also likely to be correlated with each other. This complex set of correlations make it difficult to tease apart the individual responses. These correlations can, however, be dealt with using statistical control by framing the multiple regression model in the context of a structural equation model (Hoyle 1995). A structural equation model (often referred to as path analysis or covariance structure analysis) can be specified as a system of multiple regression equations that partitions interactions among multiple dependent and independent variables (see the Methods section for details). In this paper we present a model that partitions the response of  $A_{\text{area}}$  to elevated CO<sub>2</sub> into independent and interactive components representing changes in an index of CO2 limitation, an index of biochemical capacity and an index of leaf morphology. Although the model is not mechanistic, it partitions known and hypothesized relationships in a conceptually and mechanistically plausible framework based on the general concepts of the PAC model. We used this linear model to re-analyse existing data from 10 C<sub>3</sub> species (three pine and seven deciduous tree species) grown at ambient and elevated CO2 concentrations. One aim of this study was to develop a computationally and data efficient tool for assessing the relative importance of changes in biochemical capacity and leaf morphology that could be applied to biogeochemical or biogeographic models of global change. We also compared responses across species to identify whether generalizations could be made from these data based on functional or taxonomic relatedness.

## MATERIALS AND METHODS

## Data

Data used in this analysis were obtained from 10 separate chamber-based elevated CO<sub>2</sub> experiments, conducted in the field, and included three pine and seven deciduous tree species (see caption to Fig. 2 for details). Data consisted of rates of net photosynthetic carbon assimilation  $[A_{area}]$  $\mu$ mol (CO<sub>2</sub>) m<sup>-2</sup> (leaf) s<sup>-1</sup>] measured at light saturation under growth conditions; leaf N per unit leaf area  $[N_{area}]$ g (N)  $m^{-2}$  (leaf)]; and leaf mass per unit area [LMA, g (leaf) m<sup>-2</sup> (leaf)]. In most data sets leaf N was determined using the same leaves that photosynthesis was measured on, although in some cases adjacent leaves were collected for N analysis. Variation in leaf N resulted from either fertilization treatments, sun versus shade leaves, leaf developmental stage, natural variation within leaf classes, or variation due to CO<sub>2</sub> treatment (see citations for details). Photosynthesis measurements were made at ecologically relevant temperatures for each species (20 to 30 °C depending on species) and measurements for single species were usually controlled to within  $\pm 2$  °C. Ambient CO<sub>2</sub> concentration was either 350 or 360  $\mu$ mol mol<sup>-1</sup> and the elevated concentration was either 650 or 700  $\mu$ mol mol<sup>-1</sup> (see citations for details).

## Structural equation model

Structural equation models are a class of general linear models that include analysis of variance (ANOVA), multiple

regression, path analysis and factor analysis (Hoyle 1995). In this analysis we used a system of multiple regression equations (presented below) to approximate known and hypothesized relationships among CO2, Aarea, Narea and LMA. Path analysis (also known as covariance structure analysis) could be used in this situation but we chose the multiple regression approach for the following reasons. First, path analysis is designed to analyse hypothesized relationships among measured and latent (unmeasured) variables that are inferred from a covariance matrix (Hoyle 1995). Because there are no latent variables in our model (other than the error terms), path analysis provides no benefit over multiple regression analysis in this case. Second, even though path analysis and regression analysis give identical slope coefficients for the relationships modelled in this study, path analysis does not provide information on the intercepts, which are important for comparing responses across species.

The system of regression equations used in this analysis was

$$LMA = \beta_0 + \beta_1 CO_2 + \varepsilon_i$$
  

$$N_{\text{area}} = \beta_2 + \beta_3 CO_2 + \beta 4LMA + \varepsilon_j$$
  

$$A_{\text{area}} = \beta_5 + \beta_6 CO_2 + \beta_7 N_{\text{area}} + \beta_8 LMA + \varepsilon_k$$
  
Model 1

The  $\beta$  coefficients for each equation are partial regression coefficients and give the slope of the relationship between the appropriate dependent and independent variable when the other independent variables in that equation are held constant. Two important coefficients in these equations are  $\beta_3$  (the direct effect of CO<sub>2</sub> on  $N_{\text{area}}$ ) and  $\beta_6$  (the direct effect of CO<sub>2</sub> on  $A_{\text{area}}$ ). Even though  $\beta_3$  has units of g (N) m<sup>-2</sup> (leaf), it is an index of change in N concentration ( $N_{\text{mass}}$ ) because it quantifies change in the mass of N per unit leaf area when leaf mass per unit area is held constant.  $\beta_6$  is an index of CO<sub>2</sub> limitation of photosynthesis because it quantifies the effect of CO<sub>2</sub> on  $A_{\text{area}}$  when  $N_{\text{area}}$  and *LMA* are both held constant. A graphical representation of this model is shown in Fig. 1.

The model was fitted to data from each species using least squares, and 95% confidence intervals for coefficients were estimated from 2000 bootstrap replicates using the bias corrected and accelerated method of Efron & Tibshirani (1993). Bootstrapping involves resampling with replacement from the original data to simulate multiple samples from a population. It is useful when the statistical properties of the comparison being made are not well understood, as is the case for calculating confidence intervals for the indirect effects of  $CO_2$  which are discussed below. Confidence intervals were used to assess the statistical significance of all coefficients.

Model 1 allows us to partition the effect of elevated  $CO_2$ on  $A_{area}$  into components representing a direct effect of  $CO_2$  ( $\beta_6$ ) and indirect effects consisting of interactions among  $CO_2$ ,  $N_{area}$  and LMA. For example, the indirect effect of  $CO_2$  on  $A_{area}$  due to  $CO_2$  induced changes in LMAis represented by the compound pathway  $CO_2 \rightarrow$  $LMA \rightarrow A_{area}$  (Fig. 1). In this pathway the arrows represent specific coefficients from model 1 with the first arrow



**Figure 1.** Graphical representation of model 1 showing the interactions among CO<sub>2</sub> concentration, leaf nitrogen content ( $N_{\text{area}}$ ), leaf mass per area (*LMA*) and leaf-level photosynthesis ( $A_{\text{area}}$ ). The  $\beta_i$  are coefficients in model 1. Intercepts and error terms have been excluded for simplicity.

being  $\beta_1$  and the second arrow  $\beta_8$ . The magnitude of this indirect effect is the product of these coefficients. Model 1 has three different pathways representing indirect effects of CO<sub>2</sub> on  $A_{\text{area}}$ : (1) CO<sub>2</sub>  $\rightarrow LMA \rightarrow A_{\text{area}}$ ; (2) CO<sub>2</sub>  $\rightarrow$  $LMA \rightarrow N_{\text{area}} \rightarrow A_{\text{area}}$ ; and (3) CO<sub>2</sub>  $\rightarrow N_{\text{area}} \rightarrow A_{\text{area}}$ (Fig. 1). Each indirect effect has units of  $\mu$ mol (CO<sub>2</sub>)  $m^{-2}$  (leaf) s<sup>-1</sup> and the total indirect effect of CO<sub>2</sub> on  $A_{\text{area}}$ is the sum of these three independent effects (Sokal & Rohlf 1981). Because these indirect effects have the same units and are additive they provide comparative information on the relative importance of each set of interactions on  $A_{\text{area}}$ .

In this analysis all independent variables except  $CO_2$  were random. Least squares regression is appropriate when independent variables are random as long as their frequency distributions are not functions of the regression coefficients (Neter, Wasserman & Kutner 1990; p 86). We assumed that this was the case for all data sets. Normality and homogeneity of residuals were checked using the original non-bootstrapped data. Because we expected correlations among the independent variables we paid particular attention to possible effects of multicolinearity on parameter estimates. Multicolinearity was checked using the original non-bootstrapped data but there was no evidence to suggest that it was a problem. We concluded that all original data sets showed adequate compliance with the assumptions of least squares regression.

#### Comparing responses across species

If the coefficients from model 1 were more similar within than between vegetation types (pines versus deciduous trees), then the accuracy of ecosystem and global models may be improved by incorporating the specific details of each group. We assessed the similarity of coefficients across species in two ways. First, we compared the estimated population-level frequency distributions obtained from the bootstrap replicates. If the means and 95% confidence limits for different species were similar then those species could be grouped on the premise that they had similar responses to elevated CO<sub>2</sub>. Otherwise those species were assumed to have different frequency distributions and therefore different responses.

Second, we used non-parametric and parametric clusteranalysis (Everitt 1980; Sokal & Rohlf 1981; Digby & Kempton 1987) on the coefficients to determine if species formed discrete groups. This would imply that the complete set of coefficients were more similar within than between groups. We used hierarchical cluster-analysis to identify potential groupings and then k-means cluster analysis to test whether those groups were robust. The hierarchical procedure compares the Euclidean distance between points (species) in *n*-dimensional space (where *n* is the number of coefficients in model 1) and links those points that are closest together into clusters. The k-means procedure is an optimization procedure that assigns observations to a predefined set of k groups so that the within group variance is minimized and the between group variance is maximized. We based k on the number of major groups identified by the hierarchical procedure. All coefficients were standardized to have a mean of zero and standard deviation of one, and the hierarchical clustering criterion was complete linkage using Euclidean distances.

## **RESULTS AND DISCUSSION**

Growth at elevated CO<sub>2</sub> increased net photosynthesis for nine of the 10 species in this study, although this increase was marginal for both Acer saccharum and Pinus radiata (Fig. 2a). Elevated  $CO_2$  did not have a significant effect on  $N_{\rm area}$  for the majority of species even though there was a generally consistent trend for  $N_{\text{area}}$  to be slightly lower under elevated versus ambient CO<sub>2</sub> (Fig. 2b). Alnus glutinosa was an exception because it displayed a significant increase in  $N_{\text{area}}$  under elevated CO<sub>2</sub>, whereas *P. radiata* displayed a significant reduction in  $N_{\text{area}}$  (Fig. 2b). In order to simplify the discussion of how these patterns in  $A_{\text{area}}$  and  $N_{\rm area}$  are explained by interactions among CO<sub>2</sub> concentration,  $N_{\text{mass}}$  and LMA, we focus on general responses across species. Individual species that differed from these general patterns are discussed separately. First we present results for the effects of elevated  $CO_2$  on LMA and  $N_{area}$ , then we link those results to the direct and indirect effects of CO<sub>2</sub> on  $A_{\text{area}}$ .

### Effects of elevated CO<sub>2</sub> on LMA and Narea

All species displayed a significantly positive intercept for the regression of LMA on CO<sub>2</sub> concentration (Fig. 3a) and increased CO<sub>2</sub> resulted in an increase in LMA for the majority of species (Fig. 3b). This is a common response for C<sub>3</sub> plants (Curtis 1996). The  $N_{\text{area}}$  intercept was also positive for the majority of species (Fig. 4a), but elevated CO<sub>2</sub> generally had a negative direct effect on  $N_{\text{area}}$  when LMAwas held constant (Fig. 4b). The fact that LMA was held



**Figure 2.** Mean and 95% confidence intervals for the net effect of elevated CO<sub>2</sub> on  $A_{area}$  (Panel a) and  $N_{area}$  (Panel b) calculated as the difference between elevated and ambient CO<sub>2</sub> treatments (elevated – ambient). Data citations are: *Acer rubrum* (Norby *et al.* 1997; n = 69); *Acer saccharum* (Norby *et al.* 1997; n = 72); *Alnus glutinosa* (Vogel & Curtis 1995; n = 23); *Betula pendula* (Rey & Jarvis 1998; n = 45); *Fagus sylvatica* (Forstreuter 1995; n = 87); *Liriodendron tulipifera* (Gunderson, Norby & Wullschleger 1993; n = 23); *Populus euramericana* (Curtis *et al.* 1995; n = 29); *Pinus ponderosa* (Tissue, Griffin & Ball 1999; n = 27); *Pinus radiata* (Whitehead *et al.* 1995; n = 33); *Pinus taeda* (Tissue, Thomas & Strain 1997; n = 42); Deciduous = the average for all deciduous trees, Pines = the average for all pines and All Species = the average for all species.

constant means that N concentration  $(N_{mass})$  decreased and suggests that structural or non-structural carbohydrates increased with CO<sub>2</sub> (e.g. Radoglou & Jarvis 1992; Ryle, Powell & Davidson 1992; Körner & Miglietta 1994; Schechter, Proctor & Elfving 1994; Thomas & Griffin 1994). *LMA* had a positive direct effect on  $N_{area}$  when CO<sub>2</sub> was held constant, with N accounting for approximately 1 to 2% of the increase in leaf mass (Fig. 4c). This positive effect of *LMA* on  $N_{area}$  suggests that leaves with higher *LMA* had more mesophyll tissue at a fixed CO<sub>2</sub> concentration.

The indirect pathway  $CO_2 \rightarrow LMA \rightarrow N_{area}$  [units, g (N) m<sup>-2</sup>] quantifies how  $CO_2$  induced changes in LMAaffects  $N_{area}$ . This indirect effect was not significant for *Betula pendula*, *Fagus sylvatica*, *Liriodendron tulipifera*, *Populus euramericana* and *Pinus ponderosa* (Fig. 4d). Of these five species, *F. sylvatica*, *L. tulipifera* and *P. ponderosa* 

did not display a significant effect of CO<sub>2</sub> on LMA (Fig. 3b), which explains why the indirect effect of  $CO_2$  on  $N_{area}$  was not significant for these species. In contrast, B. pendula and P. euramericana both displayed higher LMA under elevated versus ambient CO<sub>2</sub> (Fig. 3b), but the non-significant indirect effect of  $CO_2$  on  $N_{area}$  suggests that this increase in LMA was due largely to the accumulation of carbohydrates. Acer rubrum, A. saccharum, Alnus glutinosa and *Pinus taeda* all displayed positive effects of  $CO_2 \rightarrow LMA \rightarrow$  $N_{\text{area}}$  (Fig. 4d). For these four species, the CO<sub>2</sub>-induced increase in LMA appeared to be due at least in part to increased mesophyll tissue growth. However, for both A. rubrum and A. saccharum, the net effect of joint variation in  $N_{\text{mass}}$  and LMA due to elevated  $CO_2$  resulted in no overall change in Narea (Fig. 2b). Alnus glutinosa and P. taeda both tended to have higher  $N_{\text{area}}$  under elevated CO<sub>2</sub> (Fig. 2b). This was because both species displayed non-significant direct effects of  $CO_2$  on  $N_{area}$  (i.e. no reduction in  $N_{\text{mass}}$ ) (Fig. 4b) and large positive effects of CO<sub>2</sub> on LMA (Fig. 3b) and of LMA on  $N_{\text{area}}$  (Fig. 4c). These results suggest that both of these species displayed an increase in mesophyll tissue growth and little or no accumulation of carbohydrates with increasing CO<sub>2</sub> concentration.

*Pinus radiata* also differed from the majority of species because it displayed a significantly negative effect of  $CO_2$ 



**Figure 3.** Regression coefficients and 95% confidence intervals for the response of *LMA* to CO<sub>2</sub> concentration. Panel (a) is the *LMA* intercept ( $\beta_0$ ) and panel (b) is the effect of CO<sub>2</sub> on *LMA* ( $\beta_1$ ).

© 1999 Blackwell Science Ltd, Plant, Cell and Environment, 22, 1109-1119



**Figure 4.** Partial regression coefficients and 95% confidence intervals for the response of  $N_{\text{area}}$  to CO<sub>2</sub> concentration and *LMA*. Panel (a) is the  $N_{\text{area}}$  intercept ( $\beta_2$ ), panel (b) is the effect of CO<sub>2</sub> on  $N_{\text{area}}$  ( $\beta_3$ ), panel (c) is the effect of *LMA* on  $N_{\text{area}}$  ( $\beta_4$ ) and panel (d) is the indirect effect of CO<sub>2</sub>  $\rightarrow$  *LMA*  $\rightarrow$   $N_{\text{area}}$ .

on *LMA* (Fig. 3b) (see also Hocking & Meyer (1991), Coleman & Bazzaz (1992), Ryle *et al.* (1992) and Knapp *et al.* (1994) for other examples of negative effects of  $CO_2$ on *LMA*. This negative effect of  $CO_2$  on *LMA* resulted in a significantly negative indirect effect of  $CO_2$  on  $N_{area}$ (Fig. 4d). This negative indirect effect combined with a marginal reduction in  $N_{mass}$  to significantly reduce  $N_{area}$  at elevated  $CO_2$  (Fig. 2b). These results for *P. radiata* suggest that this species experienced both a reduction in  $N_{mass}$  and a reduction in mesophyll tissue under elevated  $CO_2$ .

## Effects of elevated CO<sub>2</sub> on A<sub>area</sub>

There was substantial variation across species for the  $A_{area}$ intercept, with some species displaying significantly positive values while others displayed significantly negative values (Fig. 5a). All species did, however, display a positive and significant direct effect of CO<sub>2</sub> on  $A_{area}$  (Fig. 5b). This direct effect suggests a significant reduction in the CO<sub>2</sub> limitation of photosynthesis because both  $N_{area}$  and LMA were held constant statistically. Thus leaves grown at elevated CO<sub>2</sub> had higher photosynthesis for a given  $N_{area}$  and a given LMA compared to leaves grown at ambient CO<sub>2</sub>. The direct effect of  $N_{area}$  on  $A_{area}$  was also positive for most species when CO<sub>2</sub> and LMA were held constant (Fig. 5c). This positive association between  $N_{area}$  and  $A_{area}$  is consistent with many previous studies (e.g. Field & Mooney 1986; Walters & Field 1987; Evans 1989; Reich *et al.* 1994). In contrast, *LMA* tended to have either no effect or a significant negative effect on  $A_{\text{area}}$  when CO<sub>2</sub> and  $N_{\text{area}}$  were held constant (Fig. 5d).

The indirect effect of  $CO_2 \rightarrow LMA \rightarrow A_{area}$  (Fig. 6a), which compares leaves with the same  $N_{area}$  but with LMAfree to vary with  $CO_2$  concentration, was either zero or significantly negative and corresponded with the direct effect of LMA on  $A_{area}$  discussed above. That is, species which displayed a negative and significant effect of LMA on  $A_{area}$ also displayed a negative and significant effect of  $CO_2 \rightarrow$  $LMA \rightarrow A_{area}$ . This negative effect of  $CO_2$  on  $A_{area}$  via LMA suggests the presence of a morphological mechanism of photosynthetic inhibition that is independent of any effect of  $CO_2$  on  $N_{area}$ .

The indirect effect of  $CO_2 \rightarrow LMA \rightarrow N_{area} \rightarrow A_{area}$  was not significant for most species (Fig. 6b), indicating that increased  $N_{area}$  due to greater mesophyll tissue growth under elevated  $CO_2$  was either non-existent or too small to affect  $A_{area}$ . Species that did display a positive effect of  $CO_2 \rightarrow LMA \rightarrow N_{area} \rightarrow A_{area}$  (i.e. morphological enhancement of photosynthesis) were those that also displayed a strong direct effect of  $N_{area} \rightarrow A_{area}$ . In contrast, the indirect effect of  $CO_2 \rightarrow N_{area} \rightarrow A_{area}$ , which compares leaves with the same LMA but with  $N_{area}$  free to vary with  $CO_2$ 



**Figure 5.** Partial regression coefficients and 95% confidence intervals for the response of  $A_{\text{area}}$  to CO<sub>2</sub> concentration,  $N_{\text{area}}$  and *LMA*. Panel (a) is the  $A_{\text{area}}$  intercept ( $\beta_5$ ), panel (b) is the effect of CO<sub>2</sub> on  $A_{\text{area}}$  ( $\beta_6$ ), panel (c) is the effect of  $N_{\text{area}}$  on  $A_{\text{area}}$  ( $\beta_7$ ) and panel (d) is the effect of *LMA* on  $A_{\text{area}}$  ( $\beta_8$ ).

concentration (i.e. variation in  $N_{\text{mass}}$ ), was significantly or marginally negative for most species (Fig. 6c). This negative indirect effect suggests a reduction in the biochemical capacity for photosynthesis at a given *LMA*, possibly due to the dilution of N caused by increased carbohydrate storage. The total indirect effect of CO<sub>2</sub> on  $A_{\text{area}}$  (Fig. 6d), which is the sum of the indirect effects discussed above, was not significant for all but two species (see below). This analysis suggests that in general, the enhancement of  $A_{\text{area}}$ under elevated CO<sub>2</sub> was dominated by the reduction in CO<sub>2</sub> limitation of photosynthesis. Morphological upregulation at elevated CO<sub>2</sub> did not have an appreciable effect on  $A_{\text{area}}$  for the majority of species in this study.

The total indirect effect of CO<sub>2</sub> on  $A_{area}$  discussed above was significantly negative for both *A. rubrum* and *B. pendula* (Fig. 6d). The negative response for these two species was due to a combination of morphological reduction of photosynthesis (CO<sub>2</sub>  $\rightarrow$  *LMA*  $\rightarrow$   $A_{area}$ , Fig. 6a) and a reduction in biochemical capacity (CO<sub>2</sub>  $\rightarrow$   $N_{area}$   $\rightarrow$   $A_{area}$ , Fig. 6c). For *A. rubrum*, this combination cancelled the positive effect of reduced CO<sub>2</sub> limitation (Fig. 5b) and explains why there was no net effect of CO<sub>2</sub> on photosynthesis for this species (Fig. 2a). For *B. pendula*, the negative effects of morphology and lower biochemical capacity nearly halved (44% reduction) the enhancement caused by reduced CO<sub>2</sub> limitation, resulting in an enhancement of  $A_{area}$  under elevated  $CO_2$  that was still significant (Fig. 2a), although substantially constrained.

## Comparisons across species

An important result of this analysis was the variation across species for most of the coefficients in model 1. Some of this variation may represent species differences in response to elevated CO<sub>2</sub>, although some of it may reflect differences in experimental design such as nutritional status, tree age, tree density, the seasonal timing of data collection, the temperature at which measurements were made, or to differences among experiments in the CO<sub>2</sub> concentrations used for the ambient and elevated treatments (see relevant citations for details). All experiments were, however, conducted under conditions that were considered ecologically relevant but necessarily artificial. Even though the variation between species may be reduced by using standardized experimental conditions, it may still reflect important natural variation that needs to be considered in large-scale models of plant responses to global change. Nevertheless, there seems to be little evidence to suggest that the taxonomically or functionally similar species presented in this study have comparable frequency distributions for many of these coefficients. This high degree of variation across species resulted in the averages for most coefficients being



**Figure 6.** Coefficients and 95% confidence intervals for the indirect effects of CO<sub>2</sub> on  $A_{area}$ . Panel (a) is CO<sub>2</sub>  $\rightarrow LMA \rightarrow A_{area}$ , panel (b) is CO<sub>2</sub>  $\rightarrow LMA \rightarrow N_{area} \rightarrow A_{area}$ , panel (c) is CO<sub>2</sub>  $\rightarrow N_{area} \rightarrow A_{area}$ , and panel (d) is the total indirect effect of CO<sub>2</sub> on  $A_{area}$ , i.e. the sum of the effects presented in panels (a), (b), and (c). Units for all panels are  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>.

non-significant for each vegetation type (averages are presented in each figure). The only coefficients for which these averages were significant were the *LMA* intercept (Fig. 4a), the  $N_{\text{area}}$  intercept (Fig. 3a) and the direct effect of CO<sub>2</sub> on  $A_{\text{area}}$  (Fig. 5b).

Despite the variation discussed above, the hierarchical cluster analysis identified four groups that were also confirmed by the k-means cluster analysis. Two of these contained only one species each, namely P. euramericana and P. radiata. The response of P. euramericana tended to be opposite that of the other species for several coefficients. This was most notable for the  $N_{\text{area}}$  intercept, the direct effect of  $CO_2$  on  $N_{area}$  and the direct effect of LMA on Narea (Fig. 7). Pinus radiata appeared to differ from the majority of species with respect to the LMA intercept, the effect of CO<sub>2</sub> on LMA and the direct effect of  $N_{\text{area}}$  on Aarea (Fig. 7). The third group consisted of A. glutinosa and B. pendula. These two species differed from the remaining six species by having relatively higher intercepts for  $N_{\text{area}}$ and Aarea, a relatively larger direct effect of CO2 on  $A_{\text{area}}$  and a more strongly negative effect of LMA on A<sub>area</sub> (Fig. 7). The last group included A. rubrum, A. saccharum, F. sylvatica, L. tulipifera, P. ponderosa and P. taeda (Fig. 7).

Although this analysis did identify apparently robust groupings of species, there were no clear associations based

on taxonomic or functional relatedness. In fact, the analysis showed that some taxonomically divergent species were comparable for the full set of coefficients presented in model 1. This may simply represent an artifact of the conditions used in each experiment, but we may be able to confirm or refute these patterns as additional data from newer experimental technologies such as Free Air CO<sub>2</sub> Enrichment become available. Nevertheless, it may be worthwhile conducting sensitivity analyses on models of global change by incorporating the range of variability observed here to determine if it has important effects on model predictions. Other sources of natural variation, such as temporal variation during the growing season and lageffects from previous growing seasons, may also affect the relationships modelled in this study. Understanding the temporal dynamics of these relationships may be crucial for developing accurate and robust models of plant growth in changing environments.

In summary, this analysis extended the PAC model of Luo *et al.* (1994) and suggested that the general enhancement of  $A_{\text{area}}$  under elevated CO<sub>2</sub> was dominated by a reduction in CO<sub>2</sub> limitation of photosynthesis as indicated by the direct effect of CO<sub>2</sub> on  $A_{\text{area}}$ . Contrary to the prediction of the PAC model we failed to support the hypothesis that reduction in biochemical capacity is frequently offset by increased *LMA*. Instead, we found that increased



**Figure 7.** Standardized coefficients for each group of species identified in the cluster analyses. Symbols on the *X*-axis represent each coefficient in model 1, with the dependent variable outside the brackets and the independent variable inside the brackets, e.g. LMA(CO2) is the direct effect of CO<sub>2</sub> on LMA. Group 1 = *P. euramericana*; Group 2 = *P. radiata*; Group 3 = *A. glutinosa* and *B. pendula*; Group 4 = *A. rubrum*, *A. saccharum*, *F. sylvatica*, *L. tulipifera*, *P. ponderosa* and *P. taeda*.

LMA due to elevated CO<sub>2</sub> can significantly reduce photosynthesis in some species through a mechanism that appeared to be morphologically based but independent of  $N_{\text{area}}$ . The nature of this mechanism is not clear but may include a combination of factors mentioned previously, such as a reduction in N allocation to photosynthetic versus non-photosynthetic compounds (Evans 1989), to greater allocation of biomass to structural versus photosynthetic components (Vitousek et al. 1990; Lloyd et al. 1992), to greater internal shading (Terashima & Hirosaka 1995), or to greater limitations to internal diffusion (Parkhurst 1994; Pachepsky et al. 1997). Nevertheless, this morphological reduction in photosynthesis combined additively with a reduction in biochemical capacity to significantly reduce potential photosynthetic enhancement at elevated CO<sub>2</sub> in two species.

## ACKNOWLEDGMENTS

This synthesis was supported by the US Department of Energy, grant number DE-FG02–95ER62084 to the CMEAL programme. The Electric Power Research Institute (EPRI) provided support for CMEAL meetings. Belinda Medlyn played an invaluable role in negotiating collaboration with members of the European Collaboration On  $CO_2$  Responses Applied to Forests and Trees programme (ECOCRAFT). Support for the initial organization of CMEAL was provided by the Desert Research Institute. Comments from three anonymous reviewers substantially improved an earlier version of this manuscript. CMEAL is a core project of the GCTE (Global Change and Terrestrial Ecosystems) programme.

## REFERENCES

- Coleman J.S. & Bazzaz F.A. (1992) Effects of  $CO_2$  and temperature on growth and resource use of co-occurring  $C_3$  and  $C_4$  annuals. *Ecology* **73**, 1244–1259.
- Curtis P.S. (1996) A meta-analysis of leaf gas exchange and nitrogen in trees grown under elevated carbon dioxide. *Plant, Cell and Environment* **19**, 127–137.
- Curtis P.S., Vogel C.S., Pregitzer K.S., Zak D.R. & Teeri J.A. (1995) Interacting effects of soil fertility and atmospheric CO<sub>2</sub> on leaf area growth and carbon gain physiology in *Populus X euramericana* (Dode) Guinier. *New Phytologist* **129**, 253–263.
- Digby P.G.N. & Kempton R.A. (1987) Multivariate Analysis of Ecological Communities. Chapman & Hall, London.
- Drake B.G., Gonzàlez-Meler M.A. & Long S.P. (1997) More efficient plants: a consequence of rising atmospheric CO<sub>2</sub>? Annual Review of Plant Physiology and Plant Molecular Biology 48, 609–639.
- Efron B. & Tibshirani R.J. (1993) *An Introduction to the Bootstrap.* Monographs on Statistics and Applied Probability 57. Chapman & Hall, New York.
- Evans J.R. (1989) Photosynthesis and nitrogen relationships in leaves of C<sub>3</sub> plants. *Oecologia* **78**, 9–19.
- Everitt B. (1980) Cluster Analysis, 2nd edn. Heinemann, London.
- Farquhar G.D., von Caemmerer S. & Berry J.A. (1980) A biochemical model of photosynthetic CO<sub>2</sub> assimilation in leaves of C<sub>3</sub> species. *Planta* **149**, 78–90.
- Field C. & Mooney H.A. (1986) The photosynthesis-nitrogen relationship in wild plants. In On the Economy of Plant Form and Function (ed. T.J. Givnish), pp. 25–55. Cambridge University Press, Cambridge.
- Forstreuter M. (1995) Bestandesstruktur und Netto-Photosynthese von jungen Buchen (*Fagus sylvatica* L.) unter erhöhter CO<sub>2</sub>-Konzentration. Verhandlungen der Gesellschaft für Ökologie (*Frankfurt*) Band 24, 283–292.

Gunderson C.A. & Wullschleger S.D. (1994) Photosynthetic accli-

© 1999 Blackwell Science Ltd, Plant, Cell and Environment, 22, 1109-1119

mation of forest trees to a doubling of atmospheric  $CO_2$ : a broader perspective. *Photosynthesis Research* **39**, 369–388.

- Gunderson C.A., Norby R.J. & Wullschleger S.D. (1993) Foliar gas exchange responses of two deciduous hardwoods during 3 years of growth in elevated CO<sub>2</sub>: no loss of photosynthetic enhancement. *Plant, Cell and Environment* **16**, 797–807.
- Harley P.C., Thomas R.B., Reynolds J.F. & Strain B.R. (1992) Modelling photosynthesis of cotton grown in elevated CO<sub>2</sub>. *Plant, Cell and Environment* **15**, 271–282.
- Hocking P.J. & Meyer C.P. (1991) Effects of CO<sub>2</sub> enrichment and nitrogen stress on growth, and partitioning of dry matter and nitrogen in wheat and maize. *Australian Journal of Plant Physiology* 18, 339–356.
- Hoyle R.H. (1995) The structural equation modelling approach: basic concepts and fundamental issues. In *Structural Equation Modeling: Concepts, Issues and Applications* (ed. R.H. Hoyle), pp. 5–22. Sage Publications, Thousand Oaks, CA.
- Knapp A.K., Cocke M., Hamerlynck E.P. & Owensby C.E. (1994) Effect of elevated CO<sub>2</sub> on stomatal density and distribution in a C4 grass and a C3 forb under field conditions. *Annals of Botany* 72, 595–599.
- Körner C. & Miglietta F. (1994) Long-term effects of naturally elevated CO<sub>2</sub> on Mediterranean grassland and forest trees. *Oecologia* **99**, 343–351.
- Lloyd J., Syvertsen J.P., Kriedemann P.E. & Farquhar G.D. (1992) Low conductances for CO<sub>2</sub> diffusion from stomata to the sites of carboxylation in leaves of woody species. *Plant, Cell and Environment* **15**, 873–899.
- Luo Y., Field C.B. & Mooney H.A. (1994) Predicting responses of photosynthesis and root fraction to elevated [CO<sub>2</sub>]a: interactions among carbon, nitrogen, and growth. *Plant, Cell and Environment* **17**, 1195–1204.
- Neter J., Wasserman W. & Kutner M.H. (1990) Applied Linear Statistical Models. Irwin, Homewood, Illinois.
- Norby R.J., Edwards N.T., Riggs J.S., Abner C.H., Wullschleger S.D. & Gunderson C.A. (1997) Temperature-controlled open-top chambers for global change research. *Global Change Biology* 3, 259–267.
- Pachepsky L.B. & Acock B. (1996) A model 2DLEAF of leaf gas exchange: development, validation, and ecological application. *Ecological Modelling* **93**, 1–18.
- Pachepsky L.B., Acock B., Hoffman-Benning S., Willmitzer L. & Fisahn J. (1997) Estimation of the anatomical, stomatal and biochemical components of differences in photosynthesis and transpiration of wild-type and transgenic (expressing yeast-derived invertase targeted to the vacuole) tobacco leaves. *Plant, Cell and Environment* 20, 1070–1078.
- Parkhurst D.F. (1994) Diffusion of CO<sub>2</sub> and other gases inside leaves. *New Phytologist* **126**, 449–479.
- Peterson A.G. & CMEAL participants (1999) Reconciling the apparent difference between mass- and area-based expressions of the photosynthesis–nitrogen relationship. *Oecologia* **118**, 144–150.
- Radoglou K.M. & Jarvis P.G. (1990) Effects of CO<sub>2</sub> enrichment on four poplar clones. I. Growth and leaf anatomy. *Annals of Botany* 65, 617–626.
- Radoglou K.M. & Jarvis P.G. (1992) The effects of carbon dioxide enrichment and nutrient supply on growth morphology and anatomy of *Phaseolus vulgaris* L. seedlings. *Annals of Botany* **70**, 245–256.
- Reich P.B. & Walters M.B. (1994) Photosynthesis-nitrogen relations in Amazonian tree species. II. Variation in nitrogen vis-à-vis specific leaf area influences mass- and area-based expressions. *Oecologia* **97**, 73–81.
- Reich P.B., Walters M.B. & Ellsworth D.S. (1997) From tropics to tundra: global convergence in plant functioning. *Proceedings of the National Academy of Science* **94**, 13730–13734.

- Reich P.B., Walters M.B., Ellsworth D.S. & Uhl C. (1994) Photosynthesis-nitrogen relations in Amazonian tree species. I. Patterns among species and communities. *Oecologia* 97, 62–72.
- Rey A. & Jarvis P.G. (1998) Long term photosynthetic acclimation to increased atmospheric CO<sub>2</sub> concentration in young birch trees (*Betula pendula* Roth.). *Tree Physiology* **18**, 441–450.
- Roumet C., Bel M.P., Sonie L., Jardon F. & Roy J. (1996) Growth response of grasses to elevated CO<sub>2</sub>: a physiological plurispecific analysis. *New Phytologist* **133**, 595–603.
- Ryle G.J.A., Powell C.E. & Davidson I.A. (1992) Growth of white clover, dependent on N<sub>2</sub> fixation, in elevated CO<sub>2</sub> and temperature. *Annals of Botany* **70**, 221–228.
- Sage R.F. (1990) A model describing the regulation of ribulose-1,5bisphosphate carboxylase, electron transport and triose-phosphate use in response to light intensity and CO<sub>2</sub> in C<sub>3</sub> plants. *Plant Physiology* 94, 1728–1734.
- Sage R.F., Sharkey T.D. & Seemann J.R. (1989) Acclimation of photosynthesis to elevated CO<sub>2</sub> in five C<sub>3</sub> species. *Plant Physiology* 89, 590–596.
- Schechter I., Proctor J.T.A. & Elfving D.C. (1994) Apple fruit removal and limb girdling affect fruit and leaf characteristics. *Journal of the American Society of Horticultural Science* **119**, 157–162.
- Sims D.A., Seemann J. & Luo Y. (1998) Elevated CO<sub>2</sub> concentration has independent effects on expansion rates and thickness of soybean leaves across light and nitrogen gradients. *Journal of Experimental Botany* **49**, 538–591.
- Sokal R.R. & Rohlf F.J. (1981) Biometry. Freeman, New York.
- Stitt M. (1991) Rising CO<sub>2</sub> levels and their potential significance for carbon flow in photosynthetic cells. *Plant, Cell and Environment* 14, 741–762.
- Terashima I. & Hirosaka K. (1995) Comparative ecophysiology of leaf and canopy photosynthesis. *Plant, Cell and Environment* 18, 1111–1128.
- Thomas R.B. & Griffin K.L. (1994) Direct and indirect effects of atmospheric carbon dioxide enrichment on leaf respiration of *Glycine max* (L.) Merr. *Plant Physiology* **104**, 355–361.
- Tissue D.T., Griffin K.L. & Ball J.T. (1999) Photosynthetic adjustment in field grown Ponderosa pine trees after six years exposure to elevated CO<sub>2</sub>. *Tree Physiology* **19**, 221–228.
- Tissue D.T., Thomas R.B. & Strain B.R. (1997) Atmospheric CO<sub>2</sub> enrichment increases growth and photosynthesis of *Pinus taeda*: a 4 year experiment in the field. *Plant, Cell and Environment* **20**, 1123–1134.
- Vitousek P.M., Field C.B. & Mantson P.A. (1990) Variation in foliar  $\delta^{13}$ C in Hawaiian *Metrosideros polymorpha*: a case of internal resistance? *Oecologia* **84**, 362–370.
- Vogel C.S. & Curtis P.S. (1995) Leaf gas exchange and nitrogen dynamics of N fixing, field-grown *Alnus glutinosa* under elevated atmospheric CO<sub>2</sub>. *Global Change Biology* 1, 55–61.
- Vu J.C.V., Allen L.H. Jr & Bowes G. (1989) Leaf ultrastructure, carbohydrates and protein of soybeans grown under CO<sub>2</sub> enrichment. *Environmental and Experimental Botany* 29, 141–147.
- Walters M.B. & Field C.B. (1987) Photosynthetic light acclimation in two rain forest *Piper* species with different ecological amplitudes. *Oecologia* 72, 449–459.
- Whitehead D., Hogan K.P., Rogers G.N.D., Byers J.N., Hunt J.E., Mc Seveny T.M., Hollinger D.Y., Dungan R.J., Earl W.B. & Bourke M.P. (1995) Performance of large open-top chambers for longterm field investigations of tree response to elevated carbon dioxide concentration. *Journal of Biogeography* 22, 307–313.
- Woodrow I.E. & Berry J.A. (1988) Enzymatic regulation of photosynthetic CO<sub>2</sub> fixation in C<sub>3</sub> plants. *Annual Revue of Plant Physiology and Plant Molecular Biology* **39**, 533–594.

Received 4 November 1998; received in revised form 5 March 1999; accepted for publication 5 March 1999

## APPENDIX

## CO<sub>2</sub> Models/Experiments Activity for improved Links (CMEAL)

CMEAL is a collaborative project aimed at improving the representation of the  $CO_2$  responses in ecosystem and global models. Participants are: C. B. Field (Carnegie Institution of Washington, Co-PI); J. T. Ball (Desert Research Institute, Co-PI); J. S. Amthor (Lawrence Livermore National Laboratory); B. Drake (Smithsonian Environmental Research Center); W. R. Emanuel (University of

Virginia); D. W. Johnson (Desert Research Institute); P. J. Hanson (Oak Ridge National Laboratory); Y. Luo (Desert Research Institute); R. E. McMurtrie (University of New South Wales, Australia); R. J. Norby (Oak Ridge National Laboratory); W. C. Oechel (California State University, San Diego); C. E. Owensby (Kansas State University); W. J. Parton (Colorado State University); A. G. Peterson (Desert Research Institute); L. L. Pierce (California State University, Monterey Bay); E. B. Rastetter (Marine Biological Laboratory); A. Ruimy (Universite Paris-Sud); S. W. Running (University of Montana); and D. R. Zak (University of Michigan).