Influences of soil volume and an elevated CO_2 level on growth and CO_2 exchange for the Crassulacean acid metabolism plant *Opuntia ficus-indica*

Park S. Nobel, Muyi Cui, Patsy M. Miller and Yiqi Luo

Nobel, P. S., Cui, M., Miller, P. M. and Luo, Y. 1994. Influences of soil volume and an elevated CO₂ level on growth and CO₂ exchange for the Crassulacean acid metabolism plant *Opuntia ficus-indica*. – Physiol. Plant. 90: 173–180.

Effects of the current (38 Pa) and an elevated (74 Pa) CO₂ partial pressure on root and shoot areas, biomass accumulation and daily net CO2 exchange were determined for Opuntia ficus-indica (L.) Miller, a highly productive Crassulacean acid metabolism species cultivated worldwide. Plants were grown in environmentally controlled rooms for 18 weeks in pots of three soil volumes (2 600, 6 500 and 26 000 cm³), the smallest of which was intended to restrict root growth. For plants in the medium-sized soil volume, basal cladodes tended to be thicker and areas of main and lateral roots tended to be greater as the CO2 level was doubled. Daughter cladodes tended to be initiated sooner at the current compared with the elevated \overline{CO}_2 level but total areas were similar by 10 weeks. At 10 weeks, daily net CO2 uptake for the three soil volumes averaged 24% higher for plants growing under elevated compared with current CO₂ levels, but at 18 weeks only 3% enhancement in uptake occurred. Dry weight gain was enhanced 24% by elevated CO2 during the first 10 weeks but only 8% over 18 weeks. Increasing the soil volume 10-fold led to a greater stimulation of daily net CO2 uptake and biomass production than did doubling the CO2 level. At 18 weeks, root biomass doubled and shoot biomass nearly doubled as the soil volume was increased 10-fold; the effects of soil volume tended to be greater for elevated CO2. The amount of cladode nitrogen per unit dry weight decreased as the CO2 level was raised and increased as soil volume increased, the latter suggesting that the effects of soil volume could be due to nitrogen limitations.

 $Key words - CO_2$, Crassulacean acid metabolism, global climate change, *Opuntia ficus-indica*, productivity, root growth, shoot growth.

P. S. Nobel (corresponding author) et al., UCLA-DOE Lab., Univ. of California, Los Angeles, CA 90024-1786, USA.

Introduction

Increasing levels of atmospheric CO_2 have been documented annually at the Mauna Loa Observatory since 1958 (Keeling and Whorf 1990), and the effects of elevated CO_2 levels on photosynthesis and productivity of many C_3 and C_4 crop species have been determined (Cure and Acock 1986, Strain and Cure 1986, Drake and Leadley 1991, Lawlor and Mitchell 1991). However, relatively few studies have examined the effects of elevated CO_2 levels on species with Crassulacean acid metabolism (CAM; Idso et al. 1986, Nobel and Hartsock 1986, Sza-

rek et al. 1987, Huerta and Ting 1988, Nobel and Garcia de Cortázar 1991). Virtually nothing is known about the effects of elevated CO_2 levels on root growth of CAM plants, which in terms of numbers of species outnumber C_4 plants more than 5-fold (Nobel 1991). Moreover, certain CAM plants, such as the widely cultivated *Opuntia ficus-indica* used in the present study, can achieve productivities that exceed those of most C_3 and C_4 species (Nobel 1988, 1991). The potentially high productivity of certain CAM species, their agronomic importance for food and fodder in many arid and semi-arid regions (Russell and Felker 1987), and the high water-use effi-

Received 27 April, 1993; revised 14 September, 1993

ciency of CAM plants in general (Kluge and Ting 1978, Nobel 1988) lend importance to understanding the effects of anticipated increases in atmospheric CO_2 levels for these species.

Although elevated CO₂ levels have little effect on net CO₂ uptake for C₄ plants, most C₃ plants initially respond to elevated atmospheric CO2 levels by increased net CO2 exchange and increased biomass accumulation (Arp 1991). However, when some C₃ species are grown at elevated CO₂ levels for periods of a few weeks or months. they acclimate to the elevated levels and the initially higher net CO₂ exchange rates decline to values similar to those measured under current CO₂ levels. Feedback inhibition due to root restriction by small soil volumes can cause the observed decline in net CO₂ exchange after long-term exposure to elevated CO₂ (Arp 1991, Thomas and Strain 1991, Berntson et al. 1993), although nutrient availability can be more important for some species (McConnaughay et al. 1993). For CAM plants, the longterm response to elevated atmospheric CO₂ levels may also be affected by acclimation or by decreased sink strength due to restricted root volumes, such as for those species that have roots occurring in rock crevices or are planted in close proximity in intensive agriculture. The present study therefore measured the net CO₂ uptake and above- and belowground growth under current and elevated CO₂ levels for the most widely cultivated CAM plant, Opuntia ficus-indica (Russell and Felker 1987, Nobel 1988), over an 18-week period for three soil volumes, the smallest of which was intended to restrict root growth.

Materials and methods

Similar-sized terminal cladodes were harvested from Opuntia ficus-indica (L.) Miller (Cactaceae) growing at the Agricultural Experiment Station. University of California, Riverside. Fresh weight was measured for each cladode, and the cut surfaces were allowed to heal for 7 days before planting. To estimate initial dry weight from initial fresh weight, 15 cladodes were dried at 80°C in a forced-draft oven until no further weight change occurred (generally 3 days). Their dry weight averaged 85 ± 5 g (mean \pm sE), which was $6.06 \pm 0.14\%$ of their fresh weight. The cladodes were ranked by fresh weight and grouped into 20 sets of four, each set containing one cladode from each quartile of the ranked fresh weights. Groups of two sets were randomly assigned to three different soil volumes in rectangular pots that were 23 cm deep: 2.61 (small pots), 6.51 (medium pots) or 261 (large pots). Cladodes were placed in a 1:1 (v:v) mixture of sand and soil from the Agricultural Experiment Station, with approximately one-third of the cladode area beneath the media surface. Sets were randomly assigned to the current local CO₂ partial pressure (an average of 38 Pa) or to an elevated CO₂ level (74 ± 1 Pa).

The plants were maintained in two controlled environment rooms at the University of California, Los Ange-

les. Both rooms had day/night air temperatures of 25°C/ 20°C. Light levels of 310 µmol m⁻² s⁻¹ photosynthetic photon flux density (PPFD, 400-700 nm; measured with a LI-190S quantum sensor [LI-COR, Lincoln, NE, USA] and provided by warm white fluorescent lamps [General Electric, Schenectady, NY, USA1) and a photoneriod of 14 h led to a total daily PPFD of 16 mol m⁻² dav⁻¹. The CO₂ partial pressure in one room was maintained at 74 Pa (36 Pa above the current local value) by introducing CO₂ and was continuously monitored with a LI-COR 6200 IRGA portable photosynthesis system calibrated with gases of known composition. Cladodes and CO₂ levels were alternated between the two rooms on a biweekly basis by moving the pots and the CO₂ addition system. All plants were watered twice per week with 0.5-strength Hoagland's solution No. 1 supplemented with micronutrients (Hoagland and Arnon 1950) for the first 2 weeks and with 0.1-strength solution thereafter. The water potential in the root zone was thereby maintained above -0.15 MPa (measured with PCT-55-15 soil thermocouple psychrometers [Wescor, Logan, UT, USA]).

At 10 and 18 weeks, gas exchange was measured using the LI-COR 6200 for plants under the current as well as the elevated CO₂ levels. A 250-cm³ leaf cuvette with an acrylic cylinder (5 cm in diameter, 2 cm long) having two layers of gasket on the protruding end was placed firmly on the side of an individual cladode to measure net CO₂ uptake. Sets of plants were also periodically harvested. The projected areas of main roots and their lateral roots were separately determined with an area meter (Delta-T Devices, Cambridge, UK) and multiplied by π to obtain surface areas. Areas of basal (planted) cladodes and the daughter cladodes developing on them were also measured. Dry weights were determined for all harvested organs after they were dried at 80°C. Total nitrogen content in shoots and roots was analyzed using a micro-Kieldahl technique (Bremner 1965) with about 0.1 g of powdered samples. Nitrogen contents were determined using a M/CFA continuous flow analyzer (Orion Scientific Instruments, Pleasantville, NY, USA). Data were statistically analyzed by two-way ANOVA using CoStat (CoHort, Berkeley, CA, USA),

Results

Organ growth

After a lag period of about 2 weeks, main roots rapidly developed on the basal (planted) cladodes (Fig. 1A). The surface area of main roots increased rapidly and after 6 weeks was higher on plants in medium-sized pots under elevated compared with current CO_2 levels (P < 0.05). The number of main roots averaged 31 per basal cladode at 2 weeks, increased relatively slowly to an average of 39 at 18 weeks, and was not significantly affected by the CO_2 level or the soil volume (P > 0.05). After 6 weeks, the surface area of lateral roots was greater than that of main roots and for plants in medium-sized pots was



Fig. 1. Influence of CO₂ level on growth of main roots (A) and lateral roots (B) of *Opuntia ficus-indica* over an 18-week period. The CO₂ levels averaged 38 Pa (current; Δ) or 74 Pa (elevated; Δ). Data are means \pm sE (unless the sE was smaller than the symbol) for n =4 plants in medium-sized pots (soil volume of 6 500 cm³).

greater under elevated compared with current CO₂ levels (Fig. 1B; P < 0.01). At 18 weeks, the surface area of both main and lateral roots increased an average of 105% as soil volume per plant increased 10-fold (P < 0.001) but was significantly higher under elevated CO₂ only for plants in the medium-sized pots (P < 0.05).

Accompanying the increases in root surface area with CO_2 level and with soil volume were changes in cladode properties for plants in medium-sized pots (Fig. 2). The mean thickness of the basal cladode decreased for just over 6 weeks after planting and then increased. Basal cladode thickness after 2 weeks averaged $7 \pm 1\%$ (mean \pm sE, n=6) greater under the elevated compared with the current CO_2 level (Fig. 2A; P < 0.01).

The decrease in thickness of the basal cladodes (Fig. 2A) was accompanied by the initial growth of firstdaughter cladodes (Fig. 2B), while the subsequent increase in thickness of the basal cladodes began soon after the major expansion of these daughter cladodes. Although first-daughter cladodes tended to be initiated sooner on basal cladodes under current compared with elevated CO₂ levels, after 4 weeks their number averaged 2.8 per basal cladode and did not significantly vary with CO₂ level (P > 0.05). At 10 weeks and thereafter, the area of such first-daughter cladodes in medium-sized pots tended to be greater under the elevated compared with the current CO₂ level (P < 0.05). However, second-daughter cladodes growing on the first-daughter cladodes tended to be produced earlier and tended to have greater surface areas under current than under elevated CO_2 levels (Fig. 2C).

CO2 uptake

At 10 weeks, the time courses of net CO₂ uptake for basal and daughter cladodes were similar for plants at current and elevated CO₂ levels, and most of the CO₂ uptake occurred at night (Fig. 3). For basal cladodes, the maximal night-time CO₂ uptake rates averaged 22% higher in elevated CO₂, and their daytime CO₂ efflux rates tended to be higher under the current CO₂ levels. As pot size increased, the nocturnal increase in net CO₂ rate occurred sooner during a daily cycle for basal and for daughter cladodes under the current CO₂ level (Fig. 3A,C). Increasing the soil volume increased the maximal CO₂ uptake rate more for basal cladodes than for daughter cladodes in the current CO₂ level (P < 0.05; Fig. 3A,C), but no significant effects on the maximal CO₂ uptake rate occurred for cladodes in elevated CO₂ (Fig. 3B,D).

Daily net CO₂ uptake integrated over 24 h for basal cladodes was significantly enhanced by elevated CO₂ (P < 0.001) and by larger pot sizes (P < 0.05) at 10 weeks



Fig. 2. Influence of CO₂ level on thickness of basal cladodes (A) and growth in area of first-daughter cladodes of *O. ficus-indica* developing on the basal cladodes (B) and second-daughter cladodes developing on the first-daughter cladodes (C) over an 18-week period for current (Δ) or elevated (\triangle) CO₂ levels. Data are means ± sE for n=4 plants in medium-sized pots.



Fig. 3. Daily course of net CO₂ exchange of *O. ficus-indica* for basal cladodes (A, B) and first-daughter cladodes (C, D) at the current CO₂ level (A, C; \bigcirc , \triangle , \square) and an elevated CO₂ level (B, D; \bigcirc , \triangle , \blacksquare) at 10 weeks. Data are means \pm se (n = 4 plants) for the three indicated pot sizes (soil volumes of 2 600 [\bigcirc , \bigcirc]; 6 500 [\triangle , \triangle]; or 26 000 [\square , \blacksquare cm³]).

(Fig. 4A). For daughter cladodes, the effects of CO₂ were significant (P < 0.01), but effects of pot size were not (P > 0.05; Fig. 4B). At 18 weeks, only pot size had a significant effect on daily net CO₂ uptake of basal and daughter cladodes (P < 0.01; Fig. 4). Daily net CO₂ uptake of basal cladodes at 18 weeks for the three pot sizes averaged 25% lower in the current CO₂ level and 51% lower in elevated CO₂ than at 10 weeks (Fig. 4A). For daughter cladodes, average daily net CO₂ uptake was essentially unchanged in the current CO₂ level and decreased 15% for elevated CO₂ at 18 weeks compared with 10 weeks (Fig. 4B).

At 10 weeks, the total surface areas of all daughter cladodes were similar for the two CO₂ treatments (Fig. 2B,C) and the three pot sizes. In contrast, at 18 weeks the surface areas for plants in small, medium and large pots were 0.171 ± 0.018 , 0.186 ± 0.012 and 0.323 ± 0.021 m², respectively, for the current CO₂ level (mean \pm sE for n=4 plants); for the elevated CO₂ level, the analogous values were 0.154 ± 0.013 , 0.188 ± 0.014 and 0.303 ± 0.020 m², respectively (P > 0.05 for CO₂ level; P < 0.01 for pot size). Whole plant daily net CO₂ uptake averaged

176

for all three pot sizes was 24% higher at 10 weeks in elevated than in current CO₂ levels (P < 0.01) and was 3% higher at 18 weeks (P > 0.05; Fig. 4C). At both 10 and 18 weeks, whole plant CO₂ uptake increased significantly with pot size (P < 0.001). Compared with plants in medium pots, whole plant CO₂ uptake averaged for the two CO₂ treatments at 18 weeks was 51% lower for small pots and 90% higher for large pots (Fig. 4C).

Biomass accumulation

At 10 weeks, CO₂ concentration significantly affected biomass accumulation in basal cladodes (P < 0.01; Fig. 5A). Dry weight of basal cladodes was then 5% lower than their initial dry weight in the current CO₂ level but 3% higher at the elevated CO₂ level (cf. Fig. 2A). The effects of CO₂ level on the biomass of basal cladodes were inconsistent at 18 weeks (Fig. 5A). Pot size did not significantly affect the dry weight of basal cladodes at 10 or 18 weeks (P > 0.5). However, increasing pot size enhanced biomass accumulation in daughter cladodes significantly at 18 weeks (P < 0.01; Fig. 5B). Total dry



Fig. 4. Daily net CO_2 uptake per unit surface area of basal cladodes (A) and first-daughter cladodes (B) and daily net CO_2 uptake per plant (C) in small (S), medium (M) and large (L) pots at the current CO_2 level (open bars) and an elevated CO_2 level (hatched bars) at 10 and 18 weeks. Data are means \pm sE (n=4 plants).

weight of daughter cladodes then averaged 16% less in small pots and 107% more in large pots than in medium pots. Dry weight of daughter cladodes at 18 weeks was not significantly affected by the CO₂ level (Fig. 5B).

Doubling the CO₂ level did not significantly affect root biomass except in medium pots (P < 0.05), for which root dry weight at 10 and 18 weeks averaged 37% higher for the elevated compared with the current CO₂ level (Fig. 5C). Increasing the pot size increased root biomass significantly at 10 weeks (P < 0.05) and at 18 weeks (P < 0.001). When averaged for the two CO₂ levels, root dry weight was 31% lower for small pots and 36% higher for large pots than for medium pots at 18 weeks. The root: shoot ratios for the two CO₂ levels then averaged 29% lower for small pots and 10% higher for large pots than for medium pots (P < 0.05).

Total plant biomass production by 10 weeks was significantly greater for the elevated CO₂ than for the current CO₂ level (P < 0.05; Fig. 3D). When averaged for the three pot sizes, plant dry weight was then 24% greater for the doubled CO₂ level. At 18 weeks, the plant dry weight gain averaged 8% higher under elevated compared with current CO₂ levels (Fig. 5D). Plant biomass production by 18 weeks increased significantly with pot size (P < 0.001). Total plant dry weight gain averaged for the two CO₂ levels was then 20% lower in small pots and 87% higher in large pots than in medium pots (Fig. 3D).

The cladode nitrogen content at 18 weeks tended to increase with pot size and to decrease with CO_2 level



Fig. 5. Dry weight changes of *O. ficus-indica* for basal cladodes (A), dry weight of daughter cladodes (B), dry weight of roots (C), and total plant dry weight production (D) in small (S), medium (M) and large (L) pots at the current CO₂ level (open bars) and an elevated CO₂ level (hatched bars) at 10 and 18 weeks. Data are means \pm sE (n=4 plants).



Fig. 6. Amount of nitrogen per unit dry weight in basal and daughter cladodes at the current CO_2 level (open bars) and an elevated CO_2 level (hatched bars) at 18 weeks in small (S), medium (M) and large (L) pots. Data are means \pm sE (n=4 plants).

(Fig. 6). For basal cladodes, nitrogen increased 33% from small to large pots (P < 0.01) and decreased 6% as the CO₂ partial pressure was increased from 38 to 74 Pa (P > 0.05; Fig. 6A). For daughter cladodes, nitrogen content increased 25% as pot size increased (P < 0.005) and decreased 8% as the CO₂ level was approximately doubled (P < 0.05). The nitrogen content for the 6 conditions averaged 1.61±0.09% of dry weight for the basal cladodes and 2.33±0.11% for the daughter cladodes (P < 0.001). The nitrogen content of roots was relatively constant; mean values ranged from 0.79 to 0.84% (n = 4 roots) of dry weight for the three pot sizes and the two CO₂ levels.

Discussion

For Opuntia ficus-indica in controlled environment growth rooms, increasing the soil volume 10-fold had greater effects on CO₂ uptake and biomass production than did doubling the CO₂ level. Plant daily net CO₂ uptake at 10 weeks was enhanced 24% by raising the CO₂ partial pressure from 38 to 74 Pa. At 18 weeks such enhancement depended on soil volume and was absent for plants in small pots that restricted root growth. A similar decline in enhanced net CO₂ uptake with time of exposure to elevated CO2 has been measured for annual crops (DeLucia et al. 1985, Ehret and Jolliffe 1985) and a perennial grass (Tissue and Oechel 1987). This decline has been attributed to chloroplast damage due to excessive carbohydrate accumulation in the shoots (Farrar and Williams 1991) and to changes in ribulose-1,5-bisphosphate carboxylase-oxygenase activity (Sage et al. 1989) as well as to limitations on water or nutrient uptake by roots (Berntson et al. 1993, McConnaughay et al. 1993). A 10-fold increase in soil volume increased plant daily net CO₂ uptake by O. ficus-indica over 50% at 10 and 18 weeks. When the soil volume is increased 10-fold for C₃ species, the rate of net CO2 uptake also increases, especially for crop plants (Arp 1991). Larger soil volumes can enhance photosynthetic capacity by providing more water and nutrients and by increasing the sink strength represented by growing roots (Thomas and Strain 1991, Wong and Osmond 1991, Berntson et al. 1993). Thus, the greater sink strength of roots at the doubled CO_2 level, as evidenced by the greater area for main and lateral roots of *O. ficus indica*, probably is related to its greater net CO_2 uptake at the higher CO_2 level.

Initially, basal cladodes of O, ficus-indica decreased in thickness, as their biomass was utilized for the growth of roots and daughter cladodes. At 10 weeks, basal cladodes at the current CO₂ level had a net loss of dry weight. whereas a modest increase in dry weight occurred for basal cladodes exposed to the elevated CO₂ level. The latter biomass increase for basal cladodes in part reflected their greater net CO₂ uptake at the doubled CO₂ level. At 10 weeks, daughter cladodes also had greater daily net CO₂ uptake for the elevated compared with the current CO₂ level. Greater daily CO₂ uptake by the basal and daughter cladodes helped to account for the 24% greater dry weight gain at 10 weeks when the CO₂ level was doubled. By 18 weeks, the enhancement in daily CO₂ uptake caused by doubling the CO₂ level was absent for basal cladodes in small and medium pots and for daughter cladodes in small pots, as root properties apparently then became limiting. This caused the enhanced dry weight gain due to the elevated CO₂ level to average only 8% at 18 weeks.

Increasing soil volume increased the shoot dry weight of *O. ficus-indica* more at elevated than at the current CO_2 levels, similar to results for cotton (Thomas and Strain 1991). The average root:shoot dry-weight ratio for *O. ficus-indica* was slightly higher under elevated compared with the current CO_2 level, consistent with the average 2% increase in root:shoot ratio found for C_3 species as the CO_2 level doubles (Arp 1991, Chu et al. 1992, Rogers et al. 1992). The surface area of both main and lateral roots of *O. ficus-indica* was substantially greater at the doubled CO_2 level for plants in mediumsized pots. Because increasing the pot size 10-fold approximately doubled root growth for *O. ficus-indica* in both current and elevated CO_2 levels, its root:shoot ratio substantially increased with increases in soil volume.

Increases in soil volume, which led to increases in root surface area, could lead to greater uptake of water, nitrogen and other nutrients. Because the soil was kept wet $(\psi_{soil} > -0.15 \text{ MPa})$ under all conditions, nutrients may have been the limiting factor. Indeed, the amount of nitrogen per unit dry weight in both basal and daughter cladodes of *O. ficus-indica* increased as pot size increased. Coupled with the increase in dry weight with pot size, the total amount of nitrogen in the cladodes at 18 weeks approximately doubled as the soil volume was increased 10-fold. The amount of nitrogen per unit dry weight in the cladodes also tended to decrease as the CO₂ level was doubled, in agreement with results on other species (Coleman et al. 1991, Chu et al. 1992, Silvola and Ahlholm 1992).

Biomass production in response to elevated CO₂ levels varies with CAM species. The dry weight of wellwatered Agave vilmoriniana is unaffected when the CO₂ level is doubled, but as watering is decreased the fractional increase is substantial under elevated CO₂ levels (Idso et al. 1986). The dry weight of well-watered Agave deserti and Ferocactus acanthodes is about 30% greater over a 1-year period when the CO₂ level is doubled (Nobel and Hartsock 1986). Dry mass of new cladodes of O, ficus-indica growing for 26 weeks in a 1.5:1 (v:v) gravel:vermiculite mixture in relatively small pots (2 300 cm³) but watered three times per week with 0.5-strength Hoagland's solution is 23% greater when the CO₂ level is doubled (Nobel and Garcia de Cortázar 1991). The 1.5fold greater frequency of watering with a 5-fold higher concentration of nutrients may account for the greater enhancement by elevated CO2 at the same total daily PPFD compared with the present study.

As the atmospheric CO_2 level rises, increases in the biomass productivity of the agronomically important CAM species O. ficus-indica may be minimal for closely spaced plants growing at low light levels. The present study involving doubling of the CO2 level was also conducted under well-watered conditions. Under conditions of higher PPFD and limited soil moisture, which characterize the habitats of most CAM species, the responses of O. ficus-indica to elevated CO2 may be different. Nevertheless, restricted soil volume can still be expected to diminish the responses of this CAM plant to elevated atmospheric levels of CO2.

Acknowledgements - This research was supported by the Environmental Sciences Division, Office of Health and Environmental Research, U.S. Department of Energy Carbon Dioxide Research Program grant DE-FG03-91-ER61252 and contract DE-FC03-87-ER60615.

References

- Arp, W.J. 1991, Effects of source-sink relations on photosynthetic acclimation to elevated CO2. - Plant Cell Environ. 14: 869-875
- Berntson, G.M., McConnaughay, K.D.M. & Bazzaz, F.A. 1993. Elevated CO2 alters deployment of roots in "small" growth containers. - Oecologia (In press).
- Bremner, J. M. 1965. Total nitrogen -- In Methods of Soil Analysis, part 2. Chemical and Microbiological Properties (C.A. Black, D. D. Evans, J. L. White, L. E. Ensminger, F. E. Clark and R. C. Dinaur, eds), pp. 1149-1178. American Society of Agronomy, Madison, W1.
- Chu, C. C., Coleman, J. S. & Mooney, H. A. 1992. Controls of biomass partitioning between roots and shoots: Atmospheric CO2 enrichment and the acquisition and allocation of carbon and nitrogen in wild radish. - Oecologia 89: 580-587.
- Coleman, J. S., Rochefort, L., Bazzaz, F. A. & Woodward, F.I. 1991. Atmospheric CO2, plant nitrogen status and the susceptibility of plants to an acute increase in temperature. -Plant Cell Environ. 14: 667-674.
- Cure, J. D. & Acock, B. 1986. Crop responses to carbon dioxide doubling: a literature survey. - Agric. For. Meteor. 38: 127-145.
- DeLucia, E.H., Sasek, T.W. & Strain, B.R. 1985. Photosynthetic inhibition after long-term exposure to elevated levels

of atmospheric carbon dioxide. - Photosyn. Res. 7: 175-184.

- Drake, B. G. & Leadley, P. W. 1991. Canopy photosynthesis of crops and native plant communities exposed to long-term elevated CO2. - Plant Cell Environ. 14: 853-860.
- Ehret, D.L. & Jolliffe, P.A. 1985. Photosynthetic carbon dioxide exchange of bean plants grown at elevated carbon dioxide concentrations. - Can. J. Bot. 63: 2026-2030,
- Farrar, J.F. & Williams, M.L. 1991. The effects of increased atmospheric carbon dioxide and temperature on carbon partitioning, source-sink relations and respiration. - Plant Cell Environ. 14: 819-830.
- Hoagland, D. R. & Arnon, D. I. 1950. The water-culture method for growing plants without soil. - Calif. Agric. Exp. Stn Circ. 34: 1-32.
- Huerta, A. J. & Ting, I. P. 1988. Effects of various levels of CO2 on the induction of Crassulacean acid metabolism in Portulacaria afra (L.) Jacq. - Plant Physiol. 88: 183-188.
- Idso, S. B., Kimball, B. A., Anderson, M. G. & Szarek, S. R. 1986. Growth response of a succulent plant, Agave vilmoriniana, to elevated CO2. - Plant Physiol. 80: 796-797.
- Keeling, C. D. & Whorf, T. P. 1990. Atmospheric CO2 concentrations, Mauna Loa. - In TRENDS '90: A Compendium of Data on Global Change (T. A. Boden, P. Kanciruk and M. P. Farrell, eds), pp. 8-9. Oak Ridge National Laboratory, Oak Ridge, TN. ORNL/CDIAC-36.
- Kluge, M. & Ting, I.P. 1978. Crassulacean Acid Metabolism. Analysis of an Ecological Adaptation. - Springer-Verlag, Berlin. pp. 153-169. ISBN 3-540-089-9.
- Lawlor, D. W. & Mitchell, R. A. C. 1991. The effects of increasing CO2 on crop photosynthesis and productivity: a review of field studies. - Plant Cell Environ. 14: 807-818. McConnaughay, K.D.M., Berntson, G.M. & Bazzaz, F.A.
- 1993. Limitations to CO2-induced growth enhancement in pot studies. - Oecologia (In press).
- Nobel, P.S. 1988. Environmental Biology of Agaves and Cacti. - Cambridge University Press, New York, NY. pp. 62-65, 201-207, 220-222. ISBN 0-521-34322-4.
- 1991, Tansley Review No. 32 Achievable productivities of certain CAM plants: basis for high values compared with C3 and C₄ plants. - New Phytol, 199: 183-205.
- & Garcia de Cortázar, V. 1991. Growth and predicted productivity of Opuntia ficus-indica for current and elevated carbon dioxide. - Agron. J. 83: 224-230.
- & Hartsock, T.L. 1986. Short-term and long-term responses of Crassulacean acid metabolism plants to elevated CO2. -Plant Physiol. 82: 604-606.
- Rogers, H. H., Peterson, C. M., McCrimmon, J. N. & Cure, J. D. 1992. Response of plant roots to elevated atmospheric carbon dioxide. - Plant Cell Environ. 15: 749-752.
- Russell, C. & Felker, P. 1987. The prickly-pears (Opuntia spp., Cactaceae): a source of human and animal food in semiarid regions. - Econ. Bot. 41: 433-445. Sage, R. F., Sharkey, T. D. & Seemann, J. R. 1989. Acclimation
- of photosynthesis to elevated CO₂ in five C₃ species. Plant Physiol, 89: 590-596.
- Silvola, J. & Ahlholm, U. 1992. Photosynthesis in willows (Salix × dasyclados) grown at different CO₂ concentrations and fertilization levels. - Oecologia 91: 208-213.
- Strain, B. R. & Cure, J. D. 1986. Direct effects of atmospheric CO₂ enrichment on plants and ecosystems: a bibliography with abstracts. - Carbon Dioxide Information Center, Oak Ridge National Laboratory, Oak Ridge, TN. pp. 1-199. ORNL/CDIC-13.
- Szarek, S. R., Holthe, P. A. & Ting, I. P. 1987. Minor physiological response to elevated CO₂ by the CAM plant Agave vilmoriniana. - Plant Physiol. 83: 938-940.
- Thomas, R. B. & Strain, B. R. 1991. Root restriction as a factor in photosynthetic acclimation of cotton seedlings grown in elevated carbon dioxide. - Plant Physiol, 96: 627-634. Tissue, D. T. & Oechel, W. C. 1987. Response of *Eriophorum*

vaginatum to elevated CO₂ and temperature in the Alaskan tussock tundra. – Ecology 68: 401–410.
Wong, S. C. & Osmond, C. B. 1991. Elevated atmospheric par-tial pressure of CO₂ and plant growth. III. Interactions be-tween *Triticum aestivum* (C₃) and *Echinochloa frumentacea*

 (C_4) during growth in mixed culture under different $CO_2,$ N-nutrition and irradiance treatments, with emphasis on below-ground responses estimated using the $\delta^{13}C$ value of root biomass. – Aust. J. Plant Physiol. 18: 137–152.

Edited by A. J. Stemler

This document is a scanned copy of a printed document. No warranty is given about the accuracy of the copy. Users should refer to the original published version of the material.